

Supplementary Material

Type	Name	Alignment	Position
I	LCC	-----SNPYQRGPNP-----	11
I	Cut190	-----MRGSHHHHHGSNPYERGPDP-----	22
TfCut2		-----ANPYERGPNP-----	11
IIb	PETase	--MNFPRASRLMQAAVLGGLMAVSA-----AATAQTNPYARGPNPT-----	39
PET12		MPPDCVLPRRLAAAALLASATLVP-----SAAATNPYQRGPDPT-----	41
IIa	PET5	--MN---KSILKKLSFGTSVLLVSMNALSWTPSPTPNPDPIDPDTPCQDDCDFTRGPNP-----	55
PE-H		MPFNK--KSVLA--LCGAGALLFSMSALANNPAP-----TDPGDGGGSAYQRGPDPS-----	49
I	LCC	RSALT-ADGPF SVAT YTV SRLS VSG FGGG VI YY PT GTS -LT FG GIAM SP GT A DASS LAW-----	69
I	Cut190	EDSIEAIRGPFSVATERVSSF-ASGFGGGTIYYPRETDEGTFGAVAVAPGFTASQGMSW-----	81
TfCut2		DALLEARSGPFSVSEENVSRLSASGFGGGTIYYPREN--NTYGAVAISPGYTGEASIAW-----	69
IIb	PETase	AASLEASAGPFTVRSFTVS-R-PSGYAGT VYY PT NAG-GTVGAIAIVPGYTARQSSIKW-----	96
PET12		TRDLED SRGPFRYASTNVR-S-PSGYAGTIYYPTDVS-GSVGAVAVVPGYLARQSSIRW-----	98
IIa	PET5	PSSLEASTGPYSVATR SVASS-VSG FGGG TLHYP T NTT-GTMGAI AVV PGFL LQESS IDF-----	113
PE-H		VSFLEADRGQYSVRSSRVSSL-VSG FGGG TIYY PT GTT-GTMGAVV VPGF VS AESS IDW-----	107
I	LCC	LGRR LASHG FV VL VINT NSRF DYP DS RAS QLSA AL NYL RT---SSPSA VRAR LDA N RL AV-----	126
I	Cut190	YGERV ASQGF IVF TID TNTR LDQ PGQ RGR QL LA ALD YL VE---RSDRK V RER LDPN RL AV-----	138
TfCut2		LGERI A SHG FV VIT IDT IT LDQ PDS RAE QL NA AL NH MIN---RAS STV RS R IDSS RL AV-----	126
IIb	PETase	WG PRLA SHG FV VIT IDT N STLD QPSS RS SQ MA AL RQ VAS LN GT SS PI YG KV DT AR MG V-----	156
PET12		WG PRLA SHG FV VIT LD TRST SDQ PAS RA SA QQ MA AL RQ V VAL SET R SS PI YG KV DP N RL AV-----	158
IIa	PET5	WG PKL A SHG FV VIT IS A NSG FD QP AS RA T QL GR ALD YV IN QNS NG NSN PIS GMV DT T RL GV-----	173
PE-H		WG PKL ASY GF VV MT I D T NT GFD QP PS RAR QIN N ALD YL VS QNS R SSS PV RG M ID TN RL GV-----	167
I	LCC	AG HSMGGG TL RIAE QN PSL KAA VPL TP WHT DKT-FN-TS VP VL I VG AE ADT VAP V S QHA-----	184
I	Cut190	MGHSMGGG SLE AT VM RPSL KASI PL TP WNL DKT-WG QV QV PTFI IGA ELD T IAP V RTH A-----	197
TfCut2		MGHSMGGG SL RL AS QRP DL KAA I PL TP WHL NKN-WSS VTV PTL II GAD LDT IAP VATH A-----	185
IIb	PETase	MGW SMGGG GL I SA A NN PSL KAA AP QAP WDS STN-FSS VTV PTL II FAC ENDS IAP VN SSA-----	215
PET12		MGW SMGGG GL I SA RD N PSL KAA VPF AP WHT N-A-FSG VQ V PTL V IAC END TVA PIS RHA-----	217
IIa	PET5	VG WSMGGG GAL QL AS-GDR LSA A IPI AP WQ NG NR FD QI ET PTL V IAC END VV VAS V N SH A-----	232
PE-H		IG WSMGGG TL RV AS-EGR KAA I PL AP WDT T SYY ASR SQAP TL I FACES D V IAP VL QHA-----	226
I	LCC	IP FYQ NLP STTP K VY VEL DM AS HF AP N-----SNNAA IS VYT IS WMK L WVD NDTRY RQFL CN-----	241
I	Cut190	KPF YE SLP S L P KAY ME LD GATH FAP N-----IP NTT I AKY V IS WL K R F V D ED TRY S QFL CP-----	254
TfCut2		KPF YN SLP S S I SKAY LE LD GATH FAP N-----IP NKI I G K Y SVA WL K R F V D NDTRY T QFL CP-----	242
IIb	PETase	KPI YD S M R-NA KQ FLE IN GG SH SC AN GS NSN Q ALI G K KV AWM K R F M D NDTRY ST F ACE-----	274
PET12		S S F YN SF S S S L A KAY LE IN GG SH FC A ND GG S I G G L L G KV Y G V S W M K R F ID ND L R Y D A F L CG-----	277
IIa	PET5	S P F YN R I P S T T D KAY LE IN GG SH FC A ND GG S I G G L L G KV Y G V S W M K R F ID ND L R Y D A F L CG-----	292
PE-H		S P F YN SLP S S I D KAF VE IN GG SH Y CG NG G SI Y ND V LS R F G V S W M K L H L D E D S R Y K Q F L CG-----	286
I	LCC	VND PALS--DFRTNNRH CQ-----	258
I	Cut190	NPT DR-----AIEE YR STC PY KLN-----	273
TfCut2		GPR DGL F--GEVEE YR STC PF YP N S S VDK LAA LE HHHHHH-----	282
IIb	PETase	NPN STRV SDF R T A-----NCS LE-----HHHHHH-----	298
PET12		A PH QAD LR S S R L S E Y R E S C P Y-----	298
IIa	PET5	P DHA A N R--S VSE YR DTC NY-----	310
PE-H		P NHT SDS--Q IS DY RG NC PY L-----EH HHHHHH-----	312

Figure S1 Multiple sequence alignment of PE-H with amino acid sequences of different cutinases and PET hydrolases using the program Clustal Omega. The enzymes were assigned to different types of polyester hydrolases. Amino acid residues of the catalytic triad are marked by a red triangle, disulfide forming cysteine residues are highlighted in orange and connected by an orange line. Amino acids of the extended loop region, specific for type II PET hydrolases, are framed in red. Abbreviations are: leaf-branch compost metagenome cutinase (LCC); *Saccharomonospora viridis* cutinase (Cut190); *Thermobifida fusca* cutinase (TfCut2); *Ideonella sakaiensis* PET hydrolase (PETase); *Polyangium brachysporum* PET hydrolase (PET12); *Oleispira antarctica* PET hydrolase (PET5).

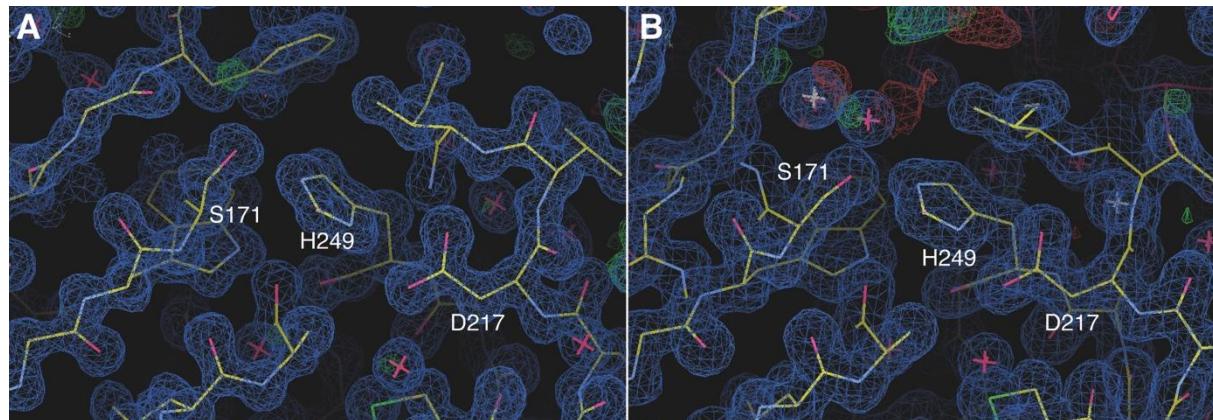


Figure S2 Electron densities around the active site of (A) PE-H WT (PDB code 6SBN) and (B) PE-H Y250S (PDB code 6SCD) with residues of the catalytic triad highlighted with labels. 2FoFc contoured at 1σ .

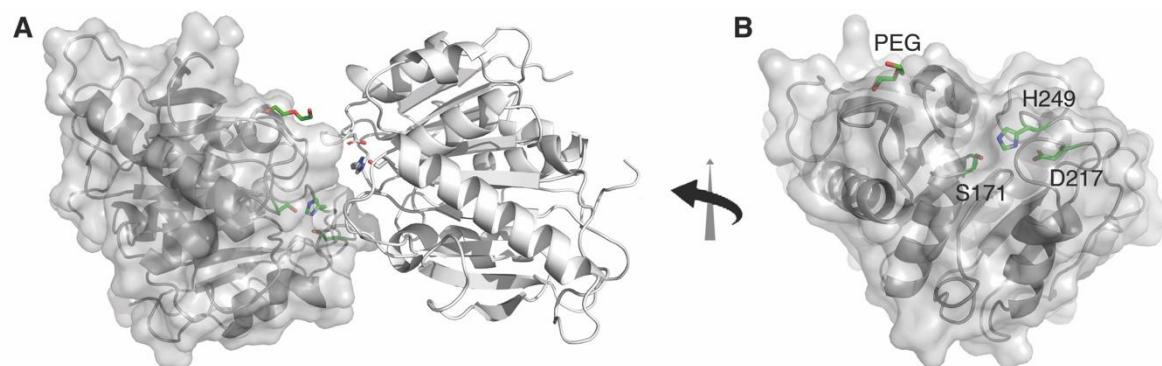


Figure S3 (A) Asymmetric unit containing two molecules of PE-H Y250S (PDB code 6SCD) depicted as cartoon showing the monomer with bound PEG molecule also in surface representation. In (B) the left molecule is rotated counter-clockwise with the residues of the catalytic triad as well as the PEG molecule shown as green sticks.

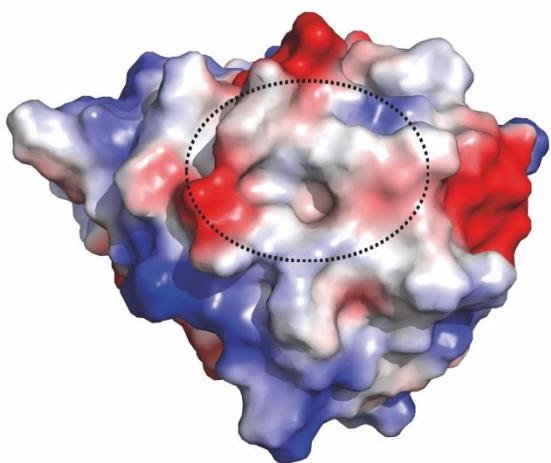
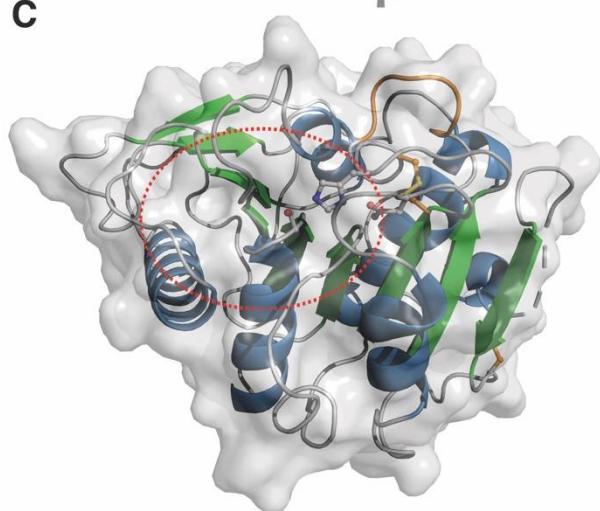
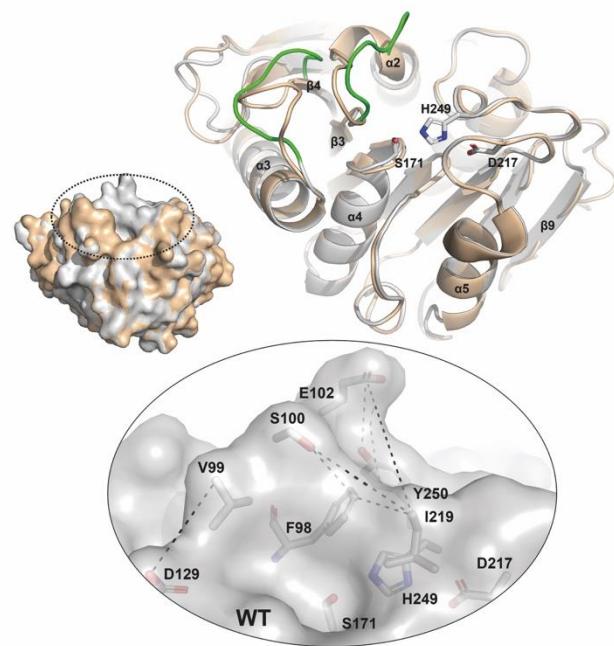
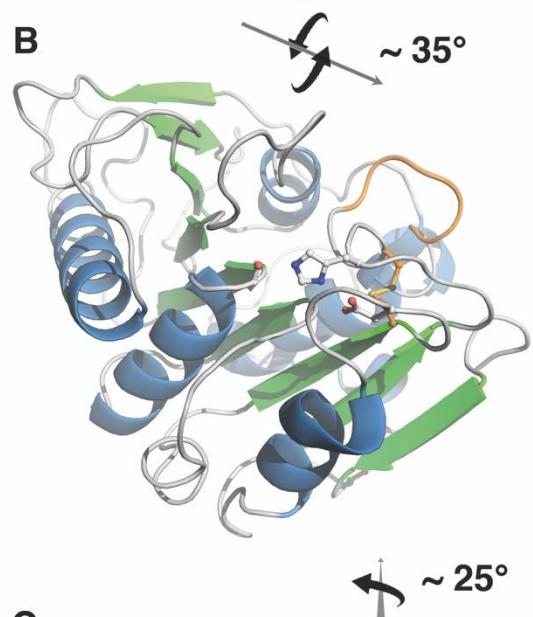
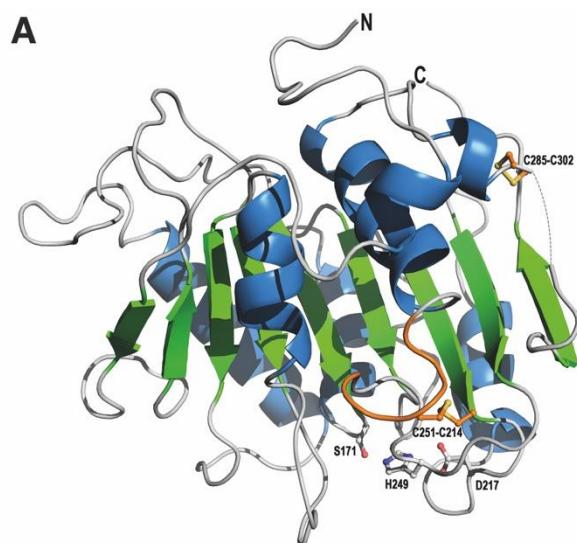


Figure S4 (A) Orientation of PE-H as reference to Figure 4. (B) PE-H is rotated for around 35° from bottom to top to zoom in the active site (as in Figure 5). In (C) PE-H is additionally rotated counter-clockwise for around 25° for a better view on the active site cleft (as in Figure S5).

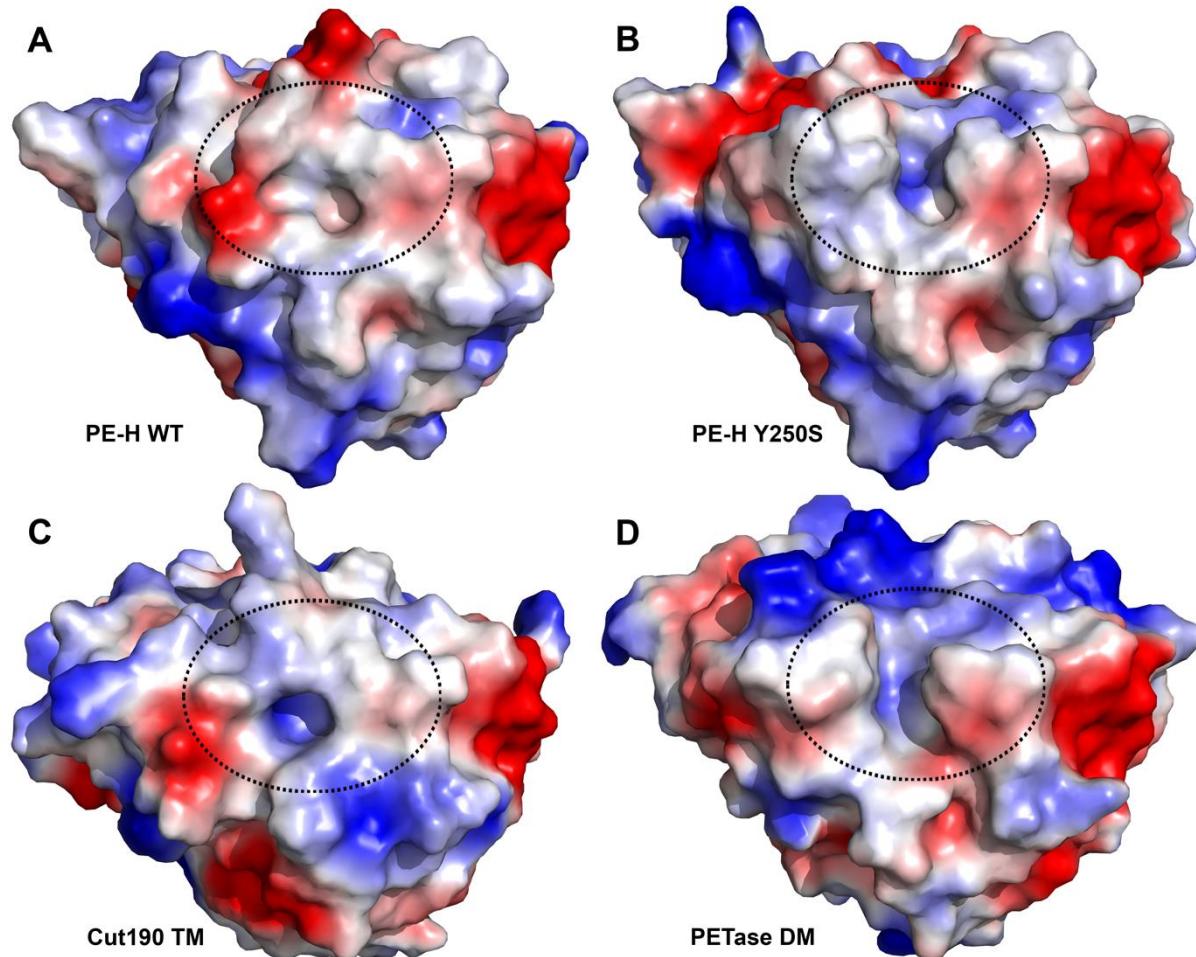


Figure S5 Surface representation of PE-H variants and two structural homologues. (A) shows wild type PE-H (PDB code 6SBN), (B) PE-H Y250S (PDB code 6SCD), (C) Cut190 triple mutant (TM) from *S. viridis* (PDB code 5ZRR) and (D) PETase double mutant (DM) from *I. sakaiensis* (PDB code 5XH3). Color code of the electrostatic surface is blue for positive, red for negative charge. All molecules are shown in identical orientation with the active site cleft positioned in the middle and marked by a dashed line.

Table S1 Amino acid composition of PETase (6EQE) and PE-H (6SBN) at selected positions as marked in the main text (i to iii).

	active site			(i)	(ii)	(iii)					
PETase	S160	D206	H237	S238	W159	S242	G243	N244	S245	N246	Q247
PE-H	S171	D217	H249	Y250	W170	G254	G255	S256	I257	Y258	N259

Table S2 Thermal melting points (Tm) of PE-H (WT) and different variants determined by nano differential scanning fluorimetry. The top row shows the respective single amino acid substitutions. Mutations G254S, S256N, I257S, Y258N, and N259Q were combined (ext.) to resemble the amino acid combination of the extended loop region of PETase.

	WT	G254S	S256N	I257S	Y258N	N259Q	ext.	Y250S
Tm [°C]	50.8	39.7	49.5	48.5	43.8	45.2	39.4	49.8

Table S3 Data collection and refinement statistics of PE-H

	PE-H	PE-H Y250S
Beamline	ID29, ESRF, Grenoble	P13, DESY, Hamburg
Crystal parameters		
Space group	C 2 2 21	I 21 21 21
Unit cell parameters:		
a, b, c (Å)	68.81, 80.01, 88.91	94.53, 98.27, 121.23
α, β, γ (°)	90, 90, 90	90, 90, 90
Data collection and processing		
Wavelength	0.9762	0.9505
Resolution (Å)	50.00 – 1.09 (1.15 – 1.09)	77.34 – 1.35 (1.42-1.35)
Total reflections	1293506 (198125)	494296 (78069)
Unique reflections	101774 (16134)	122029 (19634)
Multiplicity	12.7 (12.3)	4.1 (4.0)
Completeness (%)	99.5 (98.4)	99.11 (99.26)
I/σ(I)	16.20 (4.70)	14.98 (2.52)
Wilson B-factor (Å ²)	10.4	16.7
R-merge	0.076 (0.429)	0.043 (0.439)
R-meas	0.080 (0.448)	0.050 (0.505)
CC 1/2	0.999 (0.970)	0.998 (0.849)
Refinement statistics		
Reflections used in refinement	99673	116011
Reflections used for R-free	2057	5715
R work (%)	10.69	11.3
R free (%)	13.70	15.2
Number of non-hydrogen atoms	2401	4590
macromolecules	2028	4048
ligands	5	49
solvent	368	493
RMS (bonds)	0.027	0.026
RMS (angles)	2.206	2.430
Ramachandran plot:		
favoured (%)	96.5	97.3
Allowed (%)	2.7	2.5
outliers (%)	0.8	0.2
Average B-factor (Å ²)	14.0	22.0
Model content		
Monomers ASU	1	2
Protein residues	263 (38-285; 292-306)	532 (40-305)
Ligand	1 ACT, 1 NA	1 SO4, 3 PO4, 1 ACT, 1 PEG, 2 GOL, 3 CL, 3 NA
Waters	368	495
PDB code	6SBN	6SCD

(Statistics for the highest resolution shell are shown in parentheses)

Table S4: Structure based alignment of wild type PE-H and PE-H Y250S against pdb entries. Top 10 of the most similar structures are given with their corresponding alignment quality (Q-score), root mean square deviation (RMSD), number of aligned residues (N_{align}), their sequence identity (Seq-%), their PDB number (Target PDB), and description of the PDB entry (Protein).

Wild type PE-H (pdb-code: 6SBN)						
##	Q-score	RMSD	N_{align}	Seq-%	Target PDB	Protein
1	0.76	1.18	246	51	5zrr:A	cutinase Cut190 S176A/S226P/R228S mutant (<i>Saccharomonospora viridis</i>)
2	0.75	1.20	245	51	5xg0:B	1 PET hydrolase (<i>Ideonella sakaiensis</i>)
3	0.75	1.18	244	51	6ane:A	2 PET hydrolase (<i>Ideonella sakaiensis</i>)
4	0.75	1.24	245	51	4cg2:A	3 PET hydrolase (<i>Thermobifida fusca</i>)
5	0.75	1.25	245	51	4cg1:A	4 PET hydrolase (<i>Thermobifida fusca</i>)
6	0.74	1.22	240	53	5luj:A	5 cutinase 2 (<i>Thermobifida cellulosilytica</i>)
7	0.74	1.25	244	52	5lul:A	6 cutinase 2 R19S/R29N/A30V mutant (<i>Thermobifida cellulosilytica</i>)
8	0.74	1.32	246	52	5luk:A	7 cutinase 2 R29N/A30V mutant (<i>Thermobifida cellulosilytica</i>)
9	0.73	1.28	245	51	5lui:A	cutinase 1 (<i>Thermobifida cellulosilytica</i>)
10	0.73	1.25	242	52	5zoa:A	8 cutinase (<i>Thermobifida fusca</i>)
PE-H Y ₂₅₀ S chain A (pdb-code 6SCD)						
1	0.84	1.17	259	51	5xh3:A	9 PET hydrolase R103G/S131A mutant (HEMT complex) (<i>Ideonella sakaiensis</i>)
2	0.84	1.17	259	51	5xfz:A	10 PET hydrolase R103G/S131A mutant (<i>Ideonella sakaiensis</i>)
3	0.83	1.11	257	51	6qgc:A	11 PETase (<i>Ideonella sakaiensis</i>)
4	0.83	1.14	257	51	5xfy:A	12 PET hydrolase S131A mutant (<i>Ideonella sakaiensis</i>)
5	0.83	1.16	257	51	5xh2:A	13 PET hydrolase R103G/S131A mutant (pNP complex) (<i>Ideonella sakaiensis</i>)
6	0.83	1.10	256	52	6eqg:C	14 PET hydrolase (<i>Ideonella sakaiensis</i>)
7	0.83	1.08	253	52	4wfk:A	15 cutinase Cut190 S226P mutant (Ca(2+)-bound state) (<i>Saccharomonospora viridis</i>)
8	0.83	1.08	253	52	4wfj:A	16 cutinase Cut190 S226P mutant (Ca(2+)-bound state) (<i>Saccharomonospora viridis</i>)
9	0.83	1.08	255	52	6eqg:B	17 PET hydrolase (<i>Ideonella sakaiensis</i>)
10	0.83	1.07	255	52	6eqh:A	18 PET hydrolase (<i>Ideonella sakaiensis</i>)